Automated Immunogold Labeling of TEM Grids

Immunogold labeling (IGL) was introduced in the 1970s to enable electron microscopists to localize the constituents of a cell by attaching (electron dense) antibody-bound colloidal gold particles to antigens on specific cellular components. Here we demonstrate use of the Microscopy Innovations mPrepTM System ASP-1000 Automated Specimen Processor for this application.

Scientists at the University of Wisconsin Medical School Electron Microscope Facility are using the mPrep[™] System ASP-1000 Automated Specimen Processor to perform unattended, fully-automated IGL grid preparation with Aurion[™] blocking buffer and protocols to achieve excellent results.



ASP-1000 Automated Specimen Processor

Thin sections of resin-embedded, nematode samples (*C. elegans*) were placed on TEM grids, which were then inserted into mPrep/gTM capsules. The capsules were mounted on the ASP-1000 and processed unattended to accomplish IGL in 7.5 hours. The following Aurion protocol was utilized:

| Reagent | Incubation Time (minutes) | Rinsing Time (minutes) | Number of Rinsing Steps |
|-------------------------------------|---------------------------------|------------------------------|-------------------------------|
| Glycine | 15 | n/a | n/a |
| Aurion Blocking Solution | 15 | 5 | 3 |
| Primary Antibody | 120 | 5 | 6 |
| Secondary Antibody (gold conjugate) | 120 | 5 | 9 |
| Glutaraldehyde | 5 | 5 | 9 |



213 Air Park Rd Suite 101 Marshfield WI 54449-8626 888.302.3925 or 715.384.3292 info@microscopyinnovations.com

© 2016 Microscopy Innovations, LLC

For more information, visit www.microscopyinnovations.com

m**Prep** System

Automated Immunogold Labeling of TEM Grids



Technical Note



For more information, contact:

Thomas E. Strader, MS-Biotech tom.strader@microscopyinnovations.com

Reagents on the ASP-1000 deck were stored in 96-well sealed microplates.

Rinsing reagents were pre-dispensed into 96-well polypropylene microplates and sealed with aluminum film. Glycine, glutaraldehyde and antibodies (primary and secondary gold conjugate) were added to the microplate immediately prior to starting the pre-programmed protocol. Samples were automatically agitated by dispensing and immediately re-aspirating 40 μ L every 10 minutes during antibody incubations and every 15 seconds during rinsing steps. Upon protocol completion, transmission electron microscopy (TEM) was performed at the UW Medical School EM Facility.



A component of the endosomal sorting complex required for transport (ESCRT) machinery localizes to the limiting membrane of a multivesicular endosome in a *C. elegans* 1-cell stage embryo, as revealed by immunogold labeling (6 nm Au particles). Scale bars = 200 nm.

Acknowledgments: Special thanks to Ben August (University of Wisconsin - Madison School of Medicine and Public Health EM Facility) for assistance in adapting <u>the Aurion post-embedding</u> <u>immunogold labeling protocol</u> to an automated version and to Elisa Frankel of the Audhya lab in the UW - Madison Department of Biomolecular Chemistry for providing specimens to process and image.

Rev 3 250517